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癌細胞外泌體雙向玩弄免疫系統來助長癌症

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- 幹細胞外泌體不能胡亂使用，如用到癌症幹細胞或是發炎性的幹細胞衍生的外泌體，其對身體可能帶來疾病和癌症的風險。
- 癌細胞的外泌體會同時促進和壓抑人體的免疫系統，只是在該促進時壓抑，該壓抑時促進，以利癌細胞的入侵，轉移和擴增。
- 癌症的病患身體中已經流串著其外泌體的各種訊號因子，其免疫細胞已經受到這些因子影響而無法毒殺癌細胞。任何療法如果沒有徹底處理這些致癌訊號，一切都會是枉然，包括化放療，標靶藥物，基轉和非基轉免疫細胞療法等。
- 本人正在研發一種免疫細胞致癌訊號清洗，健康訊號重灌之免疫細胞修復技術，以恢復其對抗癌細胞的免疫力。
- 本文的專業度較深，非此領域的人士只要能掌握摘要重點即可。專業腫瘤科醫師幹則可以此論點為開端，進行其真理的探索。

前言

外泌體在細胞間的通訊中起著不可或缺的作用，並通過傳遞信號和轉移其內容而起穿梭的作用，從而在疾病的生理和病理過程的調節中起著作用（1-3）。外泌體的載物由蛋白質，脂質，DNA（mtDNA，ssDNA，dsDNA）和 RNA（mRNA，miRNA，長非編碼 RNA）組成，它們

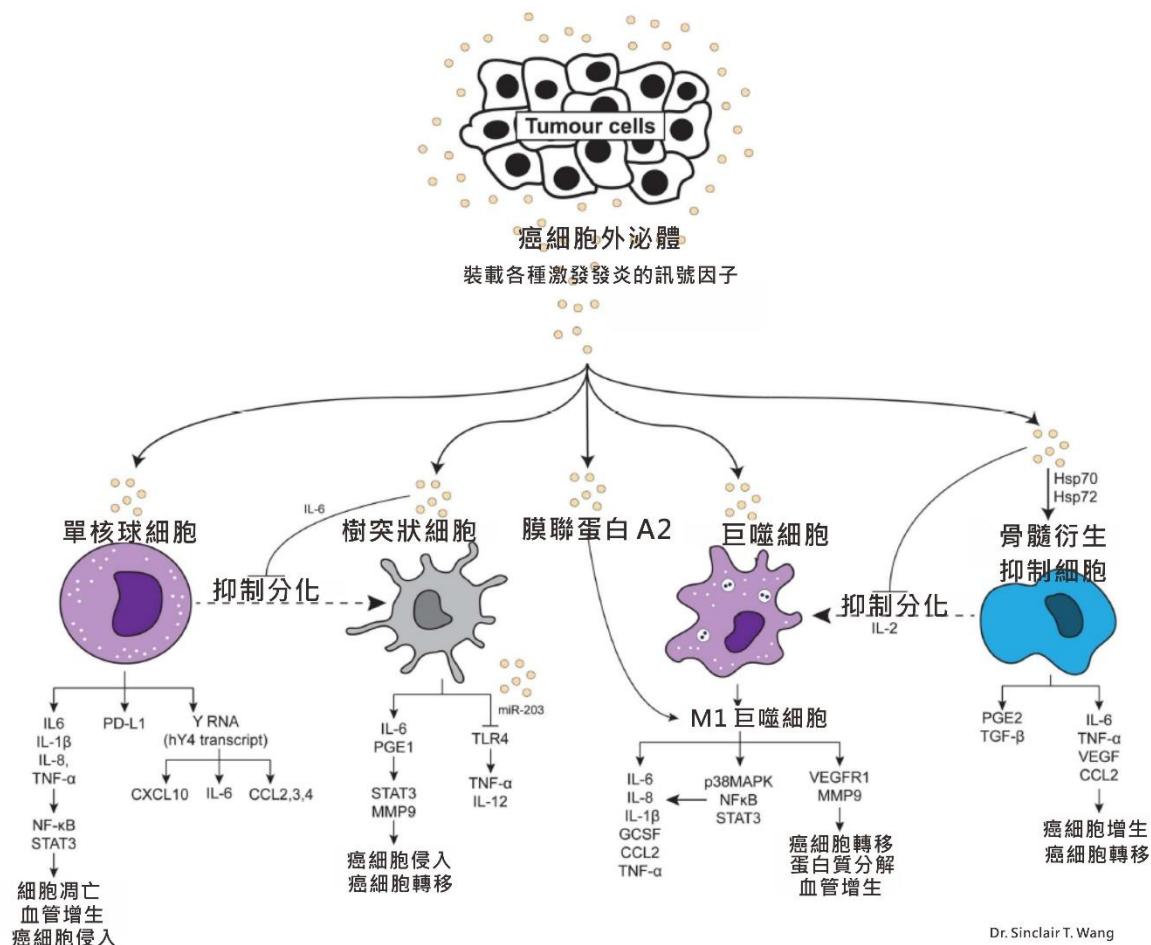


在轉移到受體細胞中時都具有某些功能（2-5）。來自腫瘤的外泌體通過其致癌成分的轉移而顯著地參與了其受體細胞的癌化。前列腺癌細胞衍生的外泌體通過運送致癌因子（例如 H-ras 和 K-ras 轉錄本）以及致癌的 miRNA miR-125b, miR-130b 和 miR-155（6）。乳腺癌患者細胞和血清的外泌體能夠以切酶依賴的方式促進健康上皮細胞形成腫瘤（7）。結腸癌的外泌體從結腸腫瘤細胞的轉移能夠在體內和體外誘導受體細胞中細胞的增殖和化學抗性的升高（8）。新血管生成的上調也受到腫瘤來源的外泌體的影響。血癌細胞衍生的外泌體能夠將 miRNA（miR-92a）轉運至內皮細胞，從而調節內皮的遷移和淋巴管的形成（11）。黑色素瘤細胞中分離出的外泌體能夠刺激癌細胞轉移性小環境的形成（10）。由於外泌體包含了其來源的細胞的所有生物活性分子，因此有可能成為癌症診斷的生物標誌（11-15）。

外泌體具有促進或抑制免疫反應的能力。源自凝血酶活化的血小板的外泌體能夠刺激造血細胞的增殖，存活和趨化性（16），並激活單核細胞釋放促炎性細胞因子並誘導 B 細胞的活化（17）。由抗原呈遞細胞，例如淋巴細胞和樹突狀細胞釋放的外泌體包含 MHC I 類分子，它們能夠潛在地介導抗原特異性 T 細胞交叉啟動（18-22）。自然殺手（NK）細胞衍生的外泌體可通過其細胞內的穿孔素和顆粒酶 B 介導抗腫瘤活性，而顆粒酶 B 對多種腫瘤細胞系具有細胞毒活性（23）。肥大細胞衍生的外泌體中表達的肽可被提呈給樹突狀細胞以刺激特異性免疫反應（24）。外泌體的各種免疫抑制作用包括通過表達死亡配體 FasL（25）和 TRAIL（26），能促進活化 T 細胞的凋亡；單核細胞對樹突狀細胞分化的損害（27）；以及抑制 NK 細胞介導的細胞毒性反應（28）。



腫瘤外泌體的促發炎作用



腫瘤外泌體對免疫分子促發炎作用的後果

腫瘤外泌體對單核細胞的促發炎作用

在慢性炎症期間，細胞因子可促進在腫瘤微環境中創造理想的生長條件，並促進腫瘤生長的進程（29-31）。腫瘤細胞及其相關的微環境可以產生改變單核細胞的募集，遷移，分化和



功能特性的分子 (32)。腫瘤細胞產生和釋放的外泌體包可以傳遞給包括單核細胞在內的受體細胞以調節其行為。在 2013 年 Bretz 等人表明，從卵巢癌患者的惡性腹水獲得的外泌體能夠調節單核細胞的生物學功能 (33)，誘導多種促炎細胞因子的產生和分泌，包括 IL-6，IL-1b 和 IL-8，以及使 TNF- α (TNF) -a 活化，隨後激活 NFkB 以及 STAT3 (33)。NFkB 的組成型激活與侵襲性表型有關，包括組織入侵和轉移以及對生長抑制的抵抗力 (34)。NFkB 和 STAT3 一起具有調節細胞凋亡，血管生成和腫瘤侵襲的能力，因此能夠抵抗免疫監視 (35)。另一研究發現 CLL 衍生的外泌體在單核細胞和巨噬細胞向腫瘤形成前表型的偏斜中起作用，並釋放腫瘤支持性細胞因子以及免疫抑制分子的表達，例如 PD-L1 來促進通過 TLR7 信號顯著增加 CCL2，CCL3，CCL4，CXCL10 和 IL-6 的分泌 (36)。

腫瘤外泌體對巨噬細胞的促發炎作用

癌細胞和腫瘤相關巨噬細胞之間通過細胞外體進行細胞間通訊，可以調節這些免疫細胞的功能和表型。乳癌 (37) 和胃癌 (38) 來源的外泌體可通過激活 NFkB 在巨噬細胞中誘導 M1 促炎反應，進而刺激包括 GCSF，IL-6，IL-8，IL-1b，CCL2 和 TNF-a。Chow 等人於 2014 年進一步揭示了 NFkB 的激活是由乳腺癌衍生的外泌體和巨噬細胞之間的相互作用介導的，並且分別受巨噬細胞和腫瘤衍生的外泌體表面上 TLR2 和棕櫚酰化蛋白配體的存在影響 (37)。在另一研究中，巨噬細胞中的 NFkB 通過將 miR-21 和 miR-29a (由腫瘤外泌體分泌) 與鼠 TLR7 和人 TLR8 結合而激活，從而觸發 TLR 介導的促轉移性炎症反應，從而促進腫瘤的生長和轉移 (39)。在乳腺癌衍生的外泌體中高表達的膜聯蛋白 A2，也在巨噬細胞介導的炎症反應中起作用 (40)。Annexin A2 通過增加 IL-6 和 TNF-a 的分泌來介導 M1 巨噬細胞活化。用含有高 Annexin A 的乳腺癌衍生外泌體引發的動物導致肺和腦部 VEGFR1 水平升高，組織中 MMP9 相應升高。腫瘤來源的外泌體上調 VEGFR1 和 MMP9 的表達分別與乳腺癌轉移，細胞外蛋白水解和血管生成有關 (39)。



腫瘤外泌體對樹突狀細胞的促發炎作用

通過抑制樹突狀細胞分化，腫瘤來源的外泌體是有效的免疫抑制劑。Yu 等人在 2007 年證明外泌體的施用引起小鼠脾臟中未分化的髓樣前體細胞的積累，並且體外將外泌體引入髓樣前體細胞導致分化的阻斷。樹突狀細胞分化的抑制作用是通過腫瘤來源的外泌體誘導 IL-6 介導的（41）。在黑色素瘤和結腸直腸癌中，腫瘤來源的外泌體能夠抑制人類單核細胞前體向樹突狀細胞的分化。這些單核細胞獲得了分泌 TGF β 的能力，從而進一步抑制了 T 淋巴細胞的增殖（42）。最近已經確定腫瘤衍生的外泌體（TEX）激活的樹突狀細胞（Ta-TEXs）能夠增加樹突狀細胞炎性介質的產生，包括 IL-6 和前列腺素 E1（PGE1）。在腫瘤來源的外泌體表面膜上發現的天然配體 Hsp105 與 TLR2 和 TLR4 的結合刺激了 IL-6 和 PGE1 的分泌。這進而通過 STAT3 的磷酸化導致腫瘤細胞侵襲和轉移的增加，進而通過與 MMP9 啟動子結合而促進基質金屬蛋白酶 9（MMP9）的轉錄（43）。此外胰腺癌衍生的外泌體通過轉移 miR-203 下調了樹突狀細胞中 TLR4 表達的表達。這導致 TNF- α 和 IL-12 的表達隨後降低（44）。

腫瘤外泌體對骨髓衍生抑制細胞（MDSC）的促發炎作用

荷瘤小鼠（45）和人類（46, 47）中 MDSC 的積累，越來越多的證據表明腫瘤微環境產生了多種抑制這些免疫調節細胞成熟和分化的因子（48-50）。通過抑制抗原加工和呈遞以及 T 細胞活化，MDSC 的積累已顯示在促進腫瘤進展中起作用，從而抑制了免疫監視和抗腫瘤免疫（51-53）。Yu 等人報導腫瘤來源的外泌體可被骨髓前體細胞吸收，並誘導這些髓樣細胞向 MDSC 途徑的分化途徑轉換（41）。腫瘤外泌體也被發現釋放促炎細胞因子 IL-6 和腫瘤生長因子 VEGF 進一步增強了其在腫瘤生長中的作用（56）。外泌體 Hsp70（57）和 Hsp72（56）均會擴展並誘導 MDSC 的激活。用外泌體 Hsp70 和 Hsp72 治療時，發現 MDSCs 顯著



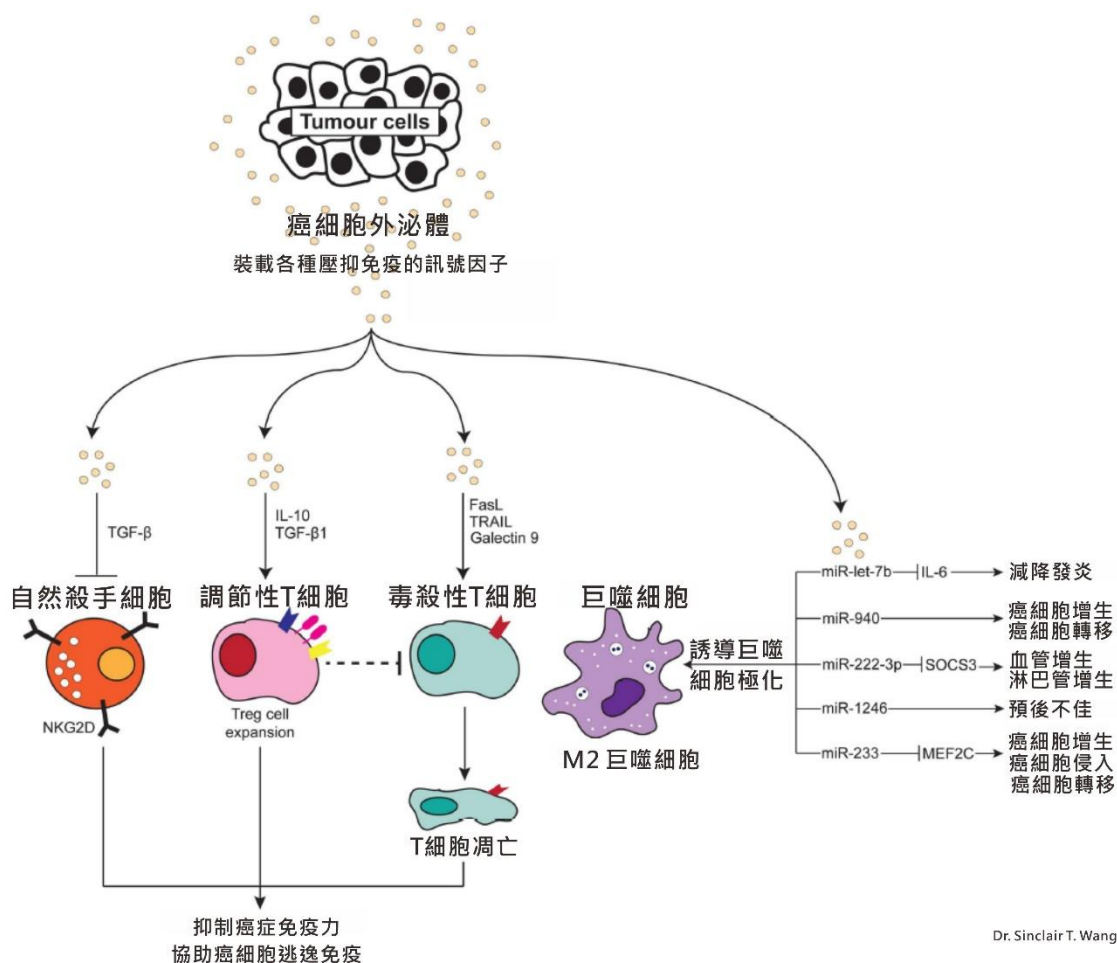
增加促炎細胞因子（包括 IL-6，TNF- α ，VEGF 和 CCL2）的產生，從而導致腫瘤生長和轉移的增加（56）。Liu 等人表明黑色素瘤外泌體對從 MyD88 基因敲除小鼠分離的骨髓（BM）前體細胞的分化沒有抑制作用，而從野生型小鼠分離的 BM 前體細胞則觀察到抑制作用。在暴露於黑色素瘤外泌體的野生型小鼠中，樹突狀細胞的百分比顯著降低，而在 MyD88 基因敲除小鼠中未觀察到顯著影響（57）。這表明 MyD88 在誘導 MDSC 中起著至關重要的作用。通過抑制巨噬細胞產生 IL-2，MDSCs 衍生的外泌體通過促進具有殺腫瘤表型的 M1 巨噬細胞向促進腫瘤的 M2 巨噬細胞的極化而促進腫瘤進展（58）。

腫瘤外泌體其他的促發炎作用

癌細胞與微環境之間的交流可以通過外泌體通過其載物的轉運來介導，這些載物包括蛋白質，DNA，信使 RNA 和微小 RNA（59, 61）。源自砷轉換的肝上皮細胞 L-02 細胞的外泌體可通過轉移外泌體 miR-155 在正常肝細胞中誘導促炎反應（62）。外泌體分泌蛋白質以會引發炎症反應，一項研究描述了從結直腸癌細胞衍生的外泌體釋放一種已知對蛋白質合成至關重要的酶賴氨酰-tRNA 合成酶（KRS），繼而誘導細胞因子和已知為 M1 和 M2 巨噬細胞標誌物的因子的釋放，包括 TNF- α ，CXC 基序趨化因子配體 10（CRG2），IL-6 和 MMP9（63）。熱休克蛋白晶體蛋白 α B（CRYAB）會因暴露於促炎細胞因子 IL-1b 和 TNF-a 而從多形膠質母細胞瘤（GBM）衍生的外泌體釋放，從而發揮抗凋亡活性（64）。此外由於輻射或疾病，GBM 細胞中的細胞因子水平增加後，GBM 衍生的外泌體蛋白質組發生了顯著變化，這可能會促進炎症的進展，腫瘤的侵襲性，血管生成和腫瘤的進展（65）。



腫瘤外泌體的抗發炎作用



腫瘤外泌體對免疫分子抗發炎作用的後果

腫瘤炎症的外泌體可通過引發免疫細胞凋亡來誘導免疫抑制（67），在愛潑斯坦-巴爾病毒（EBV）感染的鼻咽細胞中，釋放的外泌體含有高濃度的半乳凝素 9 蛋白，能夠誘導成熟的



Th1 淋巴細胞凋亡（67, 68），在大腸癌和黑色素瘤細胞中觸發活化的 CD8 + T 細胞依賴 Fas 依賴性凋亡的能力，從而有助於腫瘤從免疫系統逃逸（25，69，70）。

腫瘤外泌體對自然殺手細胞的抗發炎作用

腫瘤來源的外泌體抑制 NK 細胞的活性，作為促進癌細胞免疫逃逸的手段。這些腫瘤來源的外泌體能夠通過釋放免疫抑制性細胞因子轉化生長因子- β （TGF- β ）來阻止 NK 細胞的發育（71）。在使用同系 BALB / c 和裸鼠的研究中，源自 TS / A 或 4T.1 鼠類乳腺腫瘤細胞的外泌體能夠通過抑制 IL-2 介導的 NK 細胞活化來誘導腫瘤生長（72）。NK 細胞的增殖也受到人乳腺和黑色素瘤細胞系產生的外泌體的抑制（72）。腫瘤來源的外泌體也已顯示出誘導 Smad 磷酸化，損害細胞毒性並降低 NKG2D 受體表達，這將導致腫瘤細胞表面標誌物的丟失（73-75）。外泌體能夠通過分泌 NKG2D 配體來充當誘餌，這會下調 NKG2D 受體介導的 NK 細胞的細胞毒性（76）。

腫瘤外泌體對調節性 T 細胞（Treg）的抗發炎作用

在癌症中發現 Treg 積累在腫瘤微環境中，並通過抑制各種免疫細胞，例如 T 淋巴細胞（77），B 淋巴細胞（78），NK 細胞（79），樹突狀細胞（80）和巨噬細胞（81）。與低密度 Treg 患者相比，腫瘤基質中高密度 Treg 患者的預後也較差（82）。腫瘤來源的外泌體可通過 Treg 的間接擴增和激活來啟動免疫抑制。這將導致 Treg 抑制功能的上調並增強 Treg 對細胞凋亡的抵抗力（83，84）。Szajnik 等報導，與非癌性供體相比，骨髓瘤患者外周血中 CD24 + CD25 + FOXP3 + Treg 細胞的表達顯著增加，並且這些患者的血清中含有高濃度的腫瘤來源的外泌體。他們進一步確定，腫瘤來源的外泌體能夠通過涉及相關轉錄因子 IL-10 和 TGF- β 1 磷酸化的機制誘導 Treg 擴增（84）。



腫瘤外泌體對巨噬細胞的抗發炎作用

在肝細胞癌中，外泌體能夠間接下調促炎性細胞因子 IL-6 的表達，作為抑制免疫系統的一種手段。一項研究表明 TLR4（85、86）被發現通過 MyD88 依賴性途徑通過釋放外泌體來調節不受控制的炎症。外泌體又將 miR-let-7b 轉移至巨噬細胞以抑制促炎性 IL-6 的表達，從而減弱了腫瘤的炎症（87）。在另一研究中發現巨噬細胞受到低氧卵巢癌細胞衍生的外泌體釋放的 miR-940 的過度表達的影響，從而誘導巨噬細胞的抗炎 M2 極化，從而促進上皮性卵巢癌（EOC）細胞的增殖和遷移（88）。另一富含 EOC 來源的外泌體的 miRNA miR-222-3p 也被發現增加了 M2 巨噬細胞的極化，從而促進了腫瘤微環境中的血管生成和淋巴管生成，從而進一步促進了 EOC 的發展（89）。miR-222-3p 直接靶向 SOCS3（細胞因子信號傳導抑制劑 3），它是 JAK / STAT 通路的負調節劑（92），已被描述為控制 M1 和 M2 巨噬細胞的極化（91-93）。研究發現 SOCS3 表達的抑制與 STAT3 激活的表達增加相關（94）。

在暴露於具有功能獲得性突變體 p53 的結腸癌細胞釋放的外泌體的巨噬細胞中，也可以看到向 M2 極化的轉變。這些外泌體含有顯著高水平的 miR-1246，當轉移至鄰近的巨噬細胞時，它會刺激抗炎細胞因子和上皮間充質（EMT）促進因子的分泌增加，從而導致腫瘤形成和預後不良（95）。外泌體介導 M2 巨噬細胞極化的另一種方法是遍歷單核細胞的細胞質，以誘導肌動蛋白細胞骨架的形態變化和重組。這些膠質母細胞瘤衍生幹細胞（GSC）分泌的外泌體通過與 STAT3 結合刺激單核細胞程序性死亡配體 1（PD-L1）的增加，從而介導這種抑制性開關（94）。反過來，巨噬細胞也能夠誘導外泌體的釋放，以將 miRNA 穿梭到微環境中的相鄰細胞中。響應抗炎分子 IL-4 的激活，來自乳腺癌腫瘤的巨噬細胞已顯示出釋放含 miR-233 的外泌體的能力，該 miR-233 通過直接靶向肌細胞增強因子（Mef2c）在腫瘤的生長，侵襲和轉移中起作用（96）。



參考文獻

1. Zhang X, Yuan X, Shi H, Wu L, Qian H, Xu W. Exosomes in cancer: small particle, big player. *J Hematol Oncol.* (2015) 8:83. doi: 10.1186/s13045-015-0181-x
2. S ELA, Mager I, Breakefield XO, Wood MJ. Extracellular vesicles: biology and emerging therapeutic opportunities. *Nat Rev Drug Discov.* (2013) 12:347–57. doi: 10.1038/nrd3978
3. Kucharzewska P, Belting M. Emerging roles of extracellular vesicles in the adaptive response of tumour cells to microenvironmental stress. *J Extracell Vesicles.* (2013) 2:20304. doi: 10.3402/jev.v2i0.20304
4. Kahlert C, Melo SA, Protopopov A, Tang J, Seth S, Koch M, et al. Identification of double-stranded genomic DNA spanning all chromosomes with mutated KRAS and p53 DNA in the serum exosomes of patients with pancreatic cancer. *J Biol Chem.* (2014) 289:3869–75. doi: 10.1074/jbc.C113.53226730.
5. Valadi H, Ekstrom K, Bossios A, Sjostrand M, Lee JJ, Lotvall JO. Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat Cell Biol.* (2007) 9:654–9. doi: 10.1038/ncb1596
6. Balaj L, Lessard R, Dai L, Cho YJ, Pomeroy SL, Breakefield XO, et al. Tumour microvesicles contain retrotransposon elements and amplified oncogene sequences. *Nat Commun.* (2011) 2:180. doi: 10.1038/ncomms1180
7. Melo SA, Sugimoto H, O'Connell JT, Kato N, Villanueva A, Vidal A, et al. Cancer exosomes perform cell-independent microRNA biogenesis and promote tumorigenesis. *Cancer Cell.* (2014) 26:707–21. doi: 10.1016/j.ccell.2014.09.005
8. Soldevilla B, Rodriguez M, San Millan C, Garcia V, Fernandez-Perianez R, Gil-Calderon B, et al.



- Tumor-derived exosomes are enriched in DeltaNp73, which promotes oncogenic potential in acceptor cells and correlates with patient survival. *Hum Mol Genet.* (2014) 23:467–78. doi: 10.1093/hmg/ddt437
9. Umezu T, Ohyashiki K, Kuroda M, Ohyashiki JH. Leukemia cell to endothelial cell communication via exosomal miRNAs. *Oncogene.* (2013) 32:2747–55. doi: 10.1038/onc.2012.295
 10. Peinado H, Aleckovic M, Lavotshkin S, Matei I, Costa-Silva B, Moreno-Bueno G, et al. Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET. *Nat Med.* (2012) 18:883–91. doi: 10.1038/nm.2753
 11. Taylor DD, Gercel-Taylor C. MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer. *Gynecol Oncol.* (2008) 110:13–21. doi: 10.1016/j.ygyno.2008.04.033
 12. Gallo A, Tandon M, Alevizos I, Illei GG. The majority of microRNAs detectable in serum and saliva is concentrated in exosomes. *PLoS ONE.* (2012) 7:e30679. doi: 10.1371/journal.pone.0030679
 13. Tanaka Y, Kamohara H, Kinoshita K, Kurashige J, Ishimoto T, Iwatsuki M, et al. Clinical impact of serum exosomal microRNA-21 as a clinical biomarker in human esophageal squamous cell carcinoma. *Cancer.* (2013) 119:1159–67. doi: 10.1002/cncr.27895
 14. Li Q, Shao Y, Zhang X, Zheng T, Miao M, Qin L, et al. Plasma long noncoding RNA protected by exosomes as a potential stable biomarker for gastric cancer. *Tumour Biol.* (2015) 36:2007–12. doi: 10.1007/s13277-014-2807-y
 15. Wang J, Zhou Y, Lu J, Sun Y, Xiao H, Liu M, et al. Combined detection of serum exosomal miR-21 and HOTAIR as diagnostic and prognostic biomarkers for laryngeal squamous cell carcinoma. *Med Oncol.* (2014) 31:148. doi: 10.1007/s12032-014-0148-8



16. Baj-Krzyworzeka M, Majka M, Pratico D, Ratajczak J, Vilaire G, Kijowski J, et al. Platelet-derived microparticles stimulate proliferation, survival, adhesion, and chemotaxis of hematopoietic cells. *Exp Hematol*. (2002) 30:450–9. doi: 10.1016/S0301-472X(02)00791-9
17. Sprague DL, Elzey BD, Crist SA, Waldschmidt TJ, Jensen RJ, Ratliff TL. Platelet-mediated modulation of adaptive immunity: unique delivery of CD154 signal by platelet-derived membrane vesicles. *Blood*. (2008) 111:5028–36. doi: 10.1182/blood-2007-06-097410
18. Valenti R, Huber V, Iero M, Filipazzi P, Parmiani G, Rivoltini L. Tumorreleased microvesicles as vehicles of immunosuppression. *Cancer Res*. (2007) 67:2912–5. doi: 10.1158/0008-5472.CAN-07-0520
19. Thery C, Regnault A, Garin J, Wolfers J, Zitvogel L, Ricciardi-Castagnoli P, et al. Molecular characterization of dendritic cell-derived exosomes. Selective accumulation of the heat shock protein hsc73. *J Cell Biol*. (1999) 147:599–610. doi: 10.1083/jcb.147.3.599
20. Luketic L, Delanghe J, Sobol PT, Yang P, Frotten E, Mossman KL, et al. Antigen presentation by exosomes released from peptide-pulsed dendritic cells is not suppressed by the presence of active CTL. *J Immunol*. (2007) 179:5024–32. doi: 10.4049/jimmunol.179.8.5024
21. Admyre C, Johansson SM, Paulie S, Gabrielsson S. Direct exosome stimulation of peripheral human T cells detected by ELISPOT. *Eur J Immunol*. (2006) 36:1772–81. doi: 10.1002/eji.200535615
22. Utsugi-Kobukai S, Fujimaki H, Hotta C, Nakazawa M, Minami M. MHC class I-mediated exogenous antigen presentation by exosomes secreted from immature and mature bone marrow derived dendritic cells. *Immunol Lett*. (2003) 89:125–31. doi: 10.1016/S0165-2478(03)00128-7
23. Lugini L, Cecchetti S, Huber V, Luciani F, Macchia G, Spadaro F, et al. Immune surveillance properties of human NK cell-derived exosomes. *J Immunol*. (2012) 189:2833–42. doi:



- 10.4049/jimmunol.1101988
24. Skokos D, Botros HG, Demeure C, Morin J, Peronet R, Birkenmeier G, et al. Mast cell-derived exosomes induce phenotypic and functional maturation of dendritic cells and elicit specific immune responses in vivo. *J Immunol.* (2003) 170:3037–45. doi: 10.4049/jimmunol.170.6.3037
25. Andreola G, Rivoltini L, Castelli C, Huber V, Perego P, Deho P, et al. Induction of lymphocyte apoptosis by tumor cell secretion of FasL-bearing microvesicles. *J Exp Med.* (2002) 195:1303–16. doi: 10.1084/jem.20011624
26. Taylor DD, Gercel-Taylor C. Tumour-derived exosomes and their role in cancer-associated T-cell signalling defects. *Br J Cancer.* (2005) 92:305–11. doi: 10.1038/sj.bjc.6602316
27. Clayton A, Mason M. Exosomes in tumour immunity. *Curr Oncol.* (2009) 16:46–49. doi: 10.3747/co.v16i3.367
28. Clayton A, Mitchell JP, Court J, Mason MD, Tabi Z. Human tumor-derived exosomes selectively impair lymphocyte responses to interleukin-2. *Cancer Res.* (2007) 67:7458–66. doi: 10.1158/0008-5472.CAN-06-3456
29. Mantovani A, Muzio M, Garlanda C, Sozzani S, Allavena P. Macrophage control of inflammation: negative pathways of regulation of inflammatory cytokines. In: *Chronic Obstructive Pulmonary Disease: Pathogenesis to Treatment.* Milan (2008). doi: 10.1002/0470868678.ch8
30. Atrerkhany KN, Drutskaya MS, Nedospasov SA, Grivennikov SI, Kuprash DV. Chemokines, cytokines and exosomes help tumors to shape inflammatory microenvironment. *Pharmacol Therapeut.* (2016) 168:98–112. doi: 10.1016/j.pharmthera.2016.09.011
31. Zamarron BF, Chen WJ. Dual roles of immune cells and their factors in cancer development and progression. *Int J Biol Sci.* (2011) 7:651–8. doi: 10.7150/ijbs.7.651



32. Richards DM, Hettinger J, Feuerer M. Monocytes and macrophages in cancer: development and functions. *Cancer Microenviron.* (2013) 6:179–91. doi: 10.1007/s12307-012-0123-x
33. Bretz NP, Ridinger J, Rupp AK, Rimbach K, Keller S, Rupp C, et al. Body fluid exosomes promote secretion of inflammatory cytokines in monocytic cells via Toll-like receptor signaling. *J Biol Chem.* (2013) 288:36691–702. doi: 10.1074/jbc.M113.512806
34. Karin M, Cao Y, Greten FR, Li ZW. NF- κ B in cancer: from innocent bystander to major culprit. *Nat Rev Cancer.* (2002) 2:301–10. doi: 10.1038/nrc780
35. Bromberg J, Darnell JE Jr. The role of STATs in transcriptional control and their impact on cellular function. *Oncogene.* (2000) 19:2468–73. doi: 10.1038/sj.onc.1203476
36. Haderk F, Schulz R, Iskar M, Cid LL, Worst T, Willmund KV, et al. Tumorderived exosomes modulate PD-L1 expression in monocytes. *Sci Immunol.* (2017) 2:eaah5509. doi: 10.1126/sciimmunol.aah5509
37. Chow A, Zhou W, Liu L, Fong MY, Champer J, Van Haute D, et al. Macrophage immunomodulation by breast cancer-derived exosomes requires Toll-like receptor 2-mediated activation of NF- κ B. *Sci Rep.* (2014) 4:5750. doi: 10.1038/srep05750
38. Wu L, Zhang X, Zhang B, Shi H, Yuan X, Sun Y, et al. Exosomes derived from gastric cancer cells activate NF- κ B pathway in macrophages to promote cancer progression. *Tumour Biol.* (2016) 37:12169–80. doi: 10.1007/s13277-016-5071-5
39. Fabbri M, Paone A, Calore F, Galli R, Gaudio E, Santhanam R, et al. MicroRNAs bind to Toll-like receptors to induce prometastatic inflammatory response. *Proc Natl Acad Sci USA.* (2012) 109:E2110–6. doi: 10.1073/pnas.1209414109
40. Maji S, Chaudhary P, Akopova I, Nguyen PM, Hare RJ, Gryczynski I, et al. Exosomal annexin II promotes angiogenesis and breast cancer metastasis. *Mol Cancer Res.* (2017) 15:93–105. doi: 10.1158/1541-7786.MCR-16-0163



41. Yu S, Liu C, Su K, Wang J, Liu Y, Zhang L, et al. Tumor exosomes inhibit differentiation of bone marrow dendritic cells. *J Immunol.* (2007) 178:6867–75. doi: 10.4049/jimmunol.178.11.6867
42. Valenti R, Huber V, Filipazzi P, Pilla L, Sovenia G, Villa A, et al. Human tumor-released microvesicles promote the differentiation of myeloid cells with transforming growth factor-beta-mediated suppressive activity on T lymphocytes. *Cancer Res.* (2006) 66:9290–8. doi: 10.1158/0008-5472.CAN-06-1819
43. Shen Y, Guo D, Weng L, Wang S, Ma Z, Yang Y, et al. Tumor derived exosomes educate dendritic cells to promote tumor metastasis via HSP72/HSP105-TLR2/TLR4 pathway. *Oncoimmunology.* (2017) 6:e1362527. doi: 10.1080/2162402X.2017.1362527
44. Zhou M, Chen J, Zhou L, Chen W, Ding G, Cao L. Pancreatic cancer derived exosomes regulate the expression of TLR4 in dendritic cells via miR-203. *Cell Immunol.* (2014) 292:65–9. doi: 10.1016/j.cellimm.2014.09.004
45. David JM, Dominguez C, Hamilton DH, Palena C. The IL-8/IL-8R axis: a double agent in tumor immune resistance. *Vaccines.* (2016) 4:E22. doi: 10.3390/vaccines4030022
46. Almand B, Clark JI, Nikitina E, van Beynen J, English NR, Knight SC, et al. Increased production of immature myeloid cells in cancer patients: a mechanism of immunosuppression in cancer. *J Immunol.* (2001) 166:678–89. doi: 10.4049/jimmunol.166.1.678
47. Diaz-Montero CM, Salem ML, Nishimura MI, Garrett-Mayer E, Cole DJ, Montero AJ. Increased circulating myeloid-derived suppressor cells correlate with clinical cancer stage, metastatic tumor burden, and doxorubicin cyclophosphamide chemotherapy. *Cancer Immunol Immunother.* (2009) 58:49–59. doi: 10.1007/s00262-008-0523-4
48. Marigo I, Dolcetti L, Serafini P, Zanovello P, Bronte V. Tumor-induced tolerance and immune suppression by myeloid derived suppressor cells. *Immunol Rev.* (2008) 222:162–79. doi: 10.1111/j.1600-065X.2008.00602.x



49. Kusmartsev S, Gabrilovich DI. Inhibition of myeloid cell differentiation in cancer: the role of reactive oxygen species. *J Leukoc Biol.* (2003) 74:186–96. doi: 10.1189/jlb.0103010
50. Youn JI, Gabrilovich DI. The biology of myeloid-derived suppressor cells: the blessing and the curse of morphological and functional heterogeneity. *Eur J Immunol.* (2010) 40:2969–75. doi: 10.1002/eji.201040895
51. Whiteside TL. Exosomes and tumor-mediated immune suppression. *J Clin Invest.* (2016) 126:1216–23. doi: 10.1172/JCI81136
52. Musolino C, Allegra A, Pioggia G, Gangemi S. Immature myeloid-derived suppressor cells: a bridge between inflammation and cancer (Review). *Oncol Rep.* (2017) 37:671–83. doi: 10.3892/or.2016.5291
53. Ostrand-Rosenberg S, Sinha P. Myeloid-derived suppressor cells: linking inflammation and cancer. *J Immunol.* (2009) 182:4499–506. doi: 10.4049/jimmunol.0802740
54. Xiang X, Poliakov A, Liu C, Liu Y, Deng ZB, Wang J, et al. Induction of myeloid-derived suppressor cells by tumor exosomes. *Int J Cancer.* (2009) 124:2621–33. doi: 10.1002/ijc.24249
55. Diao J, Yang X, Song X, Chen S, He Y, Wang Q, et al. Exosomal Hsp70 mediates immunosuppressive activity of the myeloid-derived suppressor cells via phosphorylation of Stat3. *Med Oncol.* (2015) 32:453. doi: 10.1007/s12032-014-0453-2
56. Chalmin F, Ladoire S, Mignot G, Vincent J, Bruchard M, Remy-Martin JP, et al. Membrane-associated Hsp72 from tumor-derived exosomes mediates STAT3-dependent immunosuppressive function of mouse and human myeloid-derived suppressor cells. *J Clin Invest.* (2010) 120:457–71. doi: 10.1172/JCI40483
57. Burke M, Choksawangkarn W, Edwards N, Ostrand-Rosenberg S, Fenselau C. Exosomes from myeloid derived suppressor cells carry biologically active proteins. *J Proteome Res.* (2014) 13:836–43. doi: 10.1021/pr400879c



58. Abels ER. Introduction to extracellular vesicles: biogenesis, RNA cargo selection, content, release, and uptake. *CellMol Neurobiol.* (2016) 36:301–12. doi: 10.1007/s10571-016-0366-z
59. Choi DS, Kim DK, Kim YK, Gho YS. Proteomics, transcriptomics and lipidomics of exosomes and ectosomes. *Proteomics.* (2013) 13:1554–71. doi: 10.1002/pmic.201200329
60. Thakur BK, Zhang H, Becker A, Matei I, Huang Y, Costa-Silva B, et al. Double-stranded DNA in exosomes: a novel biomarker in cancer detection. *Cell Res.* (2014) 24:766–9. doi: 10.1038/cr.2014.44
61. Chen C, Luo F, Liu X, Lu L, Xu H, Yang Q, et al. NF- κ B-regulated exosomal miR-155 promotes the inflammation associated with arsenite carcinogenesis. *Cancer Lett.* (2017) 388:21–33. doi: 10.1016/j.canlet.2016.11.027
62. Jablonski KA, Gaudet AD, Amici SA, Popovich PG, Guerau-de-Arellano M. Control of the Inflammatory Macrophage Transcriptional Signature by miR-155. *PLoS ONE.* (2016) 11:e0159724. doi: 10.1371/journal.pone.0159724
63. Bala S, Marcos M, Kodys K, Csak T, Catalano D, Mandrekar P, et al. Up-regulation of microRNA-155 in macrophages contributes to increased tumor necrosis factor α (TNF α) production via increased mRNA half-life in alcoholic liver disease. *J Biol Chem.* (2011) 286:1436–44. doi: 10.1074/jbc.M110.145870
64. Kim SB, Kim HR, Park MC, Cho S, Goughnour PC, Han D, et al. Caspase-8 controls the secretion of inflammatory lysyl-tRNA synthetase in exosomes from cancer cells. *J Cell Biol.* (2017) 216:2201–16. doi: 10.1083/jcb.2016 05118
65. Kore RA, Abraham EC. Inflammatory cytokines, interleukin-1 beta and tumor necrosis factor- α , upregulated in glioblastoma multiforme, raise the levels of CRYAB in exosomes secreted by U373 glioma cells. *Biochem Biophys Res Commun.* (2014) 453:326–31. doi: 10.1016/j.bbrc.2014.09.068



66. Barros FM, Carneiro F, Machado JC, Melo SA. Exosomes and immune response in cancer: friends or foes? *Front Immunol.* (2018) 9:730. doi: 10.3389/fimmu.2018.00730
67. Klibi J, Niki T, Riedel A, Pioche-Durieu C, Souquere S, Rubinstein E, et al. Blood diffusion and Th1-suppressive effects of galectin-9-containing exosomes released by Epstein-Barr virus-infected nasopharyngeal carcinoma cells. *Blood.* (2009) 113:1957–66. doi: 10.1182/blood-2008-02-142596
68. Keryer-Bibens C, Pioche-Durieu C, Villemant C, Souquere S, Nishi N, Hirashima M, et al. Exosomes released by EBV-infected nasopharyngeal carcinoma cells convey the viral latent membrane protein 1 and the immunomodulatory protein galectin 9. *BMC Cancer.* (2006) 6:283. doi: 10.1186/1471-2407-6-283
69. Wieckowski EU, Visus C, Szajnik M, Szczepanski MJ, Storkus WJ, Whiteside TL. Tumor-derived microvesicles promote regulatory T cell expansion and induce apoptosis in tumor-reactive activated CD8+ T lymphocytes. *J Immunol.* (2009) 183:3720–30. doi: 10.4049/jimmunol.0900970
70. Huber V, Fais S, Iero M, Lugini L, Canese P, Squarcina P, et al. Human colorectal cancer cells induce T-cell death through release of proapoptotic microvesicles: role in immune escape. *Gastroenterology.* (2005) 128:1796–804. doi: 10.1053/j.gastro.2005.03.045
71. Szczepanski MJ, Szajnik M, Welsh A, Whiteside TL, Boyiadzis M. Blast-derived microvesicles in sera from patients with acute myeloid leukemia suppress natural killer cell function via membrane-associated transforming growth factor-beta1. *Haematologica.* (2011) 96:1302–9. doi: 10.3324/haematol.2010.039743
72. Liu C, Yu S, Zinn K, Wang J, Zhang L, Jia Y, et al. Murine mammary carcinoma exosomes promote tumor growth by suppression of NK cell function. *J Immunol.* (2006) 176:1375–85. doi: 10.4049/jimmunol.176.3.1375



73. Clayton A, Tabi Z. Exosomes and the MICA-NGK2D system in cancer. *Blood Cells Mol Dis.* (2005) 34:206–13. doi: 10.1016/j.bcmd.2005.03.003
74. Clayton A, Mitchell JP, Court J, Linnane S, Mason MD, Tabi Z. Human tumor-derived exosomes down-modulate NGK2D expression. *J Immunol.* (2008) 180:7249–58. doi: 10.4049/jimmunol.180.11.7249
75. Ashiru O, Boutet P, Fernandez-Messina L, Aguera-Gonzalez S, Skepper JN, Vales-Gomez M, et al. Natural killer cell cytotoxicity is suppressed by exposure to the human NGK2D ligand MICA_008 that is shed by tumor cells in exosomes. *Cancer Res.* (2010) 70:481–9. doi: 10.1158/0008-5472.CAN-09-1688
76. Hedlund M, Nagaeva O, Kargl D, Baranov V, Mincheva-Nilsson L. Thermal and oxidative stress causes enhanced release of NGK2D ligand-bearing immunosuppressive exosomes in leukemia/lymphoma T and B cells. *PLoS ONE.* (2011) 6:e16899. doi: 10.1371/journal.pone.0016899
77. Kursar M, Bonhagen K, Fensterle J, Kohler A, Hurwitz R, Kamradt T, et al. Regulatory CD4+CD25+ T cells restrict memory CD8+ T cell responses. *J Exp Med.* (2002) 196:1585–92. doi: 10.1084/jem.20011347
78. Zhao DM, Thornton AM, DiPaolo RJ, Shevach EM. Activated CD4+CD25+ T cells selectively kill B lymphocytes. *Blood.* (2006) 107:3925–32. doi: 10.1182/blood-2005-11-4502
79. Trzonkowski P, Szmit E, Mysliwska J, Dobyszyk A, Mysliwski A. CD4+CD25+ T regulatory cells inhibit cytotoxic activity of T CD8+ and NK lymphocytes in the direct cell-to-cell interaction. *Clin Immunol.* (2004) 112:258–67. doi: 10.1016/j.clim.2004.04.003
80. Misra N, Bayry J, Lacroix-Desmazes S, Kazatchkine MD, Kaveri SV. Cutting edge: human CD4+CD25+ T cells restrain the maturation and antigen presenting function of dendritic cells. *J Immunol.* (2004) 172:4676–80. doi: 10.4049/jimmunol.172.8.4676



81. Taams LS, van Amelsfort JM, Tiemessen MM, Jacobs KM, de Jong EC, Akbar AN, et al. Modulation of monocyte/macrophage function by human CD4+CD25+ regulatory T cells. *Hum Immunol.* (2005) 66:222–30. doi: 10.1016/j.humimm.2004.12.006
82. Wang Y, Ma Y, Fang Y, Wu S, Liu L, Fu D, et al. Regulatory T cell: a protection for tumour cells. *J Cell Mol Med.* (2012) 16:425–36. doi: 10.1111/j.1582-4934.2011.01437.x
83. Muller L, Simms P, Hong CS, Nishimura MI, Jackson EK, Watkins SC, et al. Human tumor-derived exosomes (TEX) regulate Treg functions via cell surface signaling rather than uptake mechanisms. *Oncoimmunology.* (2017) 6:e1261243. doi: 10.1080/2162402X.2016.1261243
84. Szajnik M, Czystowska M, Szczepanski MJ, Mandapathil M, Whiteside TL. Tumor-derived microvesicles induce, expand and up-regulate biological activities of human regulatory T cells (Treg). *PLoS ONE.* (2010) 5:e11469. doi: 10.1371/journal.pone.0011469
85. Huang B, Zhao J, Li H, He KL, Chen Y, Chen SH, et al. Toll-like receptors on tumor cells facilitate evasion of immune surveillance. *Cancer Res.* (2005) 65:5009–14. doi: 10.1158/0008-5472.CAN-05-0784
86. Kelly MG, Alvero AB, Chen R, Silasi DA, Abrahams VM, Chan S, et al. TLR-4 signaling promotes tumor growth and paclitaxel chemoresistance in ovarian cancer. *Cancer Res.* (2006) 66:3859–68. doi: 10.1158/0008-5472.CAN-05-3948
87. Li D, Jia H, Zhang H, Lv M, Liu J, Zhang Y, et al. TLR4 signaling induces the release of microparticles by tumor cells that regulate inflammatory cytokine IL-6 of macrophages via microRNA let-7b. *Oncoimmunology.* (2012) 1:687–93. doi: 10.4161/onci.19854
88. Chen X, Ying X, Wang X, Wu X, Zhu Q, Wang X. Exosomes derived from hypoxic epithelial ovarian cancer deliver microRNA-940 to induce macrophage M2 polarization. *Oncol Rep.* (2017) 38:522–8. doi: 10.3892/or.2017.5697
89. Ying X, Wu Q, Wu X, Zhu Q, Wang X, Jiang L, et al. Epithelial ovarian cancer-secreted exosomal



- miR-222-3p induces polarization of tumor-associated macrophages. *Oncotarget*. (2016) 7:43076–87. doi: 10.18632/oncotarget.9246
90. Yoshimura A, Naka T, Kubo M. SOCS proteins, cytokine signaling and immune regulation. *Nat Rev Immunol*. (2007) 7:454–65. doi: 10.1038/nri2093
91. Arnold CE, Whyte CS, Gordon P, Barker RN, Rees AJ, Wilson HM. A critical role for suppressor of cytokine signalling 3 in promoting M1 macrophage activation and function in vitro and in vivo. *Immunology*. (2014) 141:96–110. doi: 10.1111/imm.12173
92. Liu Y, Stewart KN, Bishop E, Marek CJ, Kluth DC, Rees AJ, et al. Unique expression of suppressor of cytokine signaling 3 is essential for classical macrophage activation in rodents in vitro and in vivo. *J Immunol*. (2008) 180:6270–8. doi: 10.4049/jimmunol.180.9.6270
93. Soki FN, Koh AJ, Jones JD, Kim YW, Dai J, Keller ET, et al. Polarization of prostate cancer-associated macrophages is induced by milk fat globule-EGF factor 8 (MFG-E8)-mediated efferocytosis. *J Biol Chem*. (2014) 289:24560–72. doi: 10.1074/jbc.M114.571620
94. Gabrusiewicz K, Li X, Wei J, Hashimoto Y, Marisetty AL, Ott M, et al. Glioblastoma stem cell-derived exosomes induce M2 macrophages and PDL1 expression on human monocytes. *Oncoimmunology*. (2018) 7:e1412909. doi: 10.1080/2162402X.2017.1412909
95. Cooks T, Pateras IS, Jenkins LM, Patel KM, Robles AI, Morris J, et al. Mutant p53 cancers reprogram macrophages to tumor supporting macrophages via exosomal miR-1246. *Nat Commun*. (2018) 9:771. doi: 10.1038/s41467-018-03224-w
96. Yang M, Chen J, Su F, Yu B, Su F, Lin L, et al. Microvesicles secreted by macrophages shuttle invasion-potentiating microRNAs into breast cancer cells. *Mol Cancer*. (2011) 10:117. doi: 10.1186/1476-4598-10-117